

# Human Genetics and Plant Genomics: The long and the short of it

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Simons Center for Quantitative Biology

CSHL In-House Symposium XXVI

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# Outline



1. De novo mutations in human diseases
  1. Autism Spectrum Disorder
  2. Applications to ADHD & Tourette's
2. Plant Genome Assembly
  1. Long read single molecule sequencing
  2. Other applications

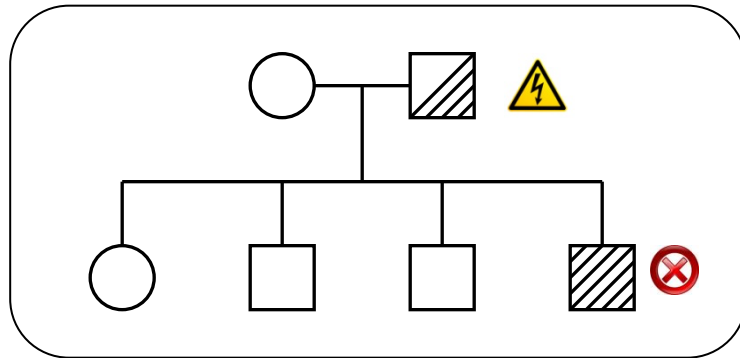
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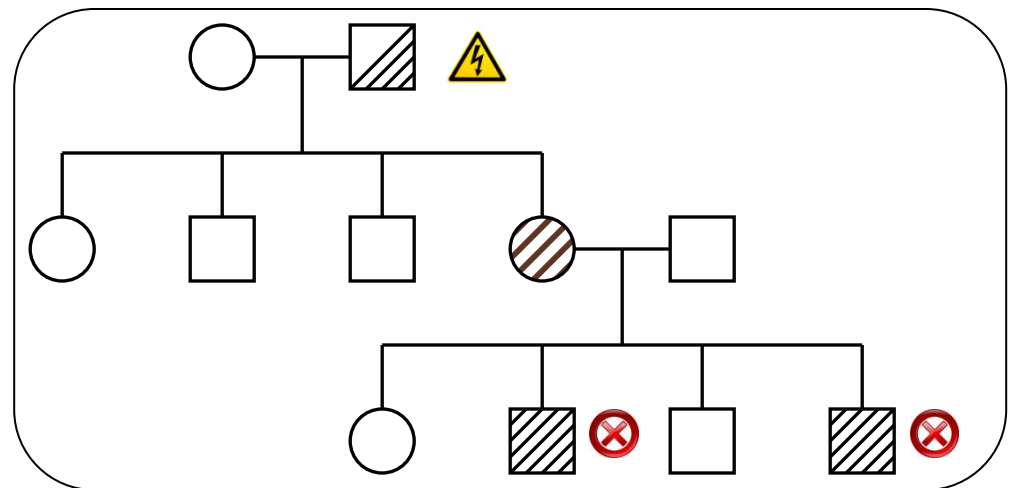
# Unified Model of Autism

## Sporadic Autism: 1 in 100



**Prediction:** De novo mutations of high penetrance contributes to autism, especially in low risk families with no history of autism.

## Familial Autism: 90% concordance in twins



### Legend



Sporadic mutation

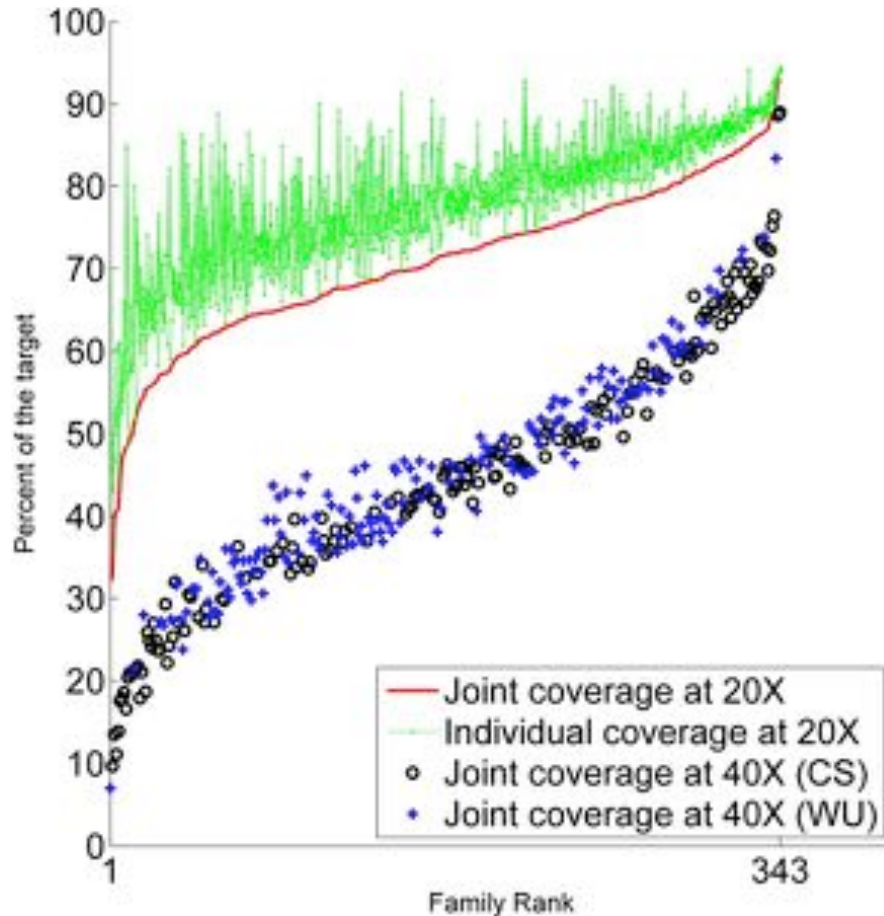


Fails to procreate

**A unified genetic theory for sporadic and inherited autism**

Zhao et al. (2007) *PNAS*. 104(31)12831-12836.

# Exome sequencing of the SSC



Sequencing of 343 families from the Simons Simplex Collection

- Parents plus one child with autism and one non-autistic sibling
- Enriched for higher-functioning individuals

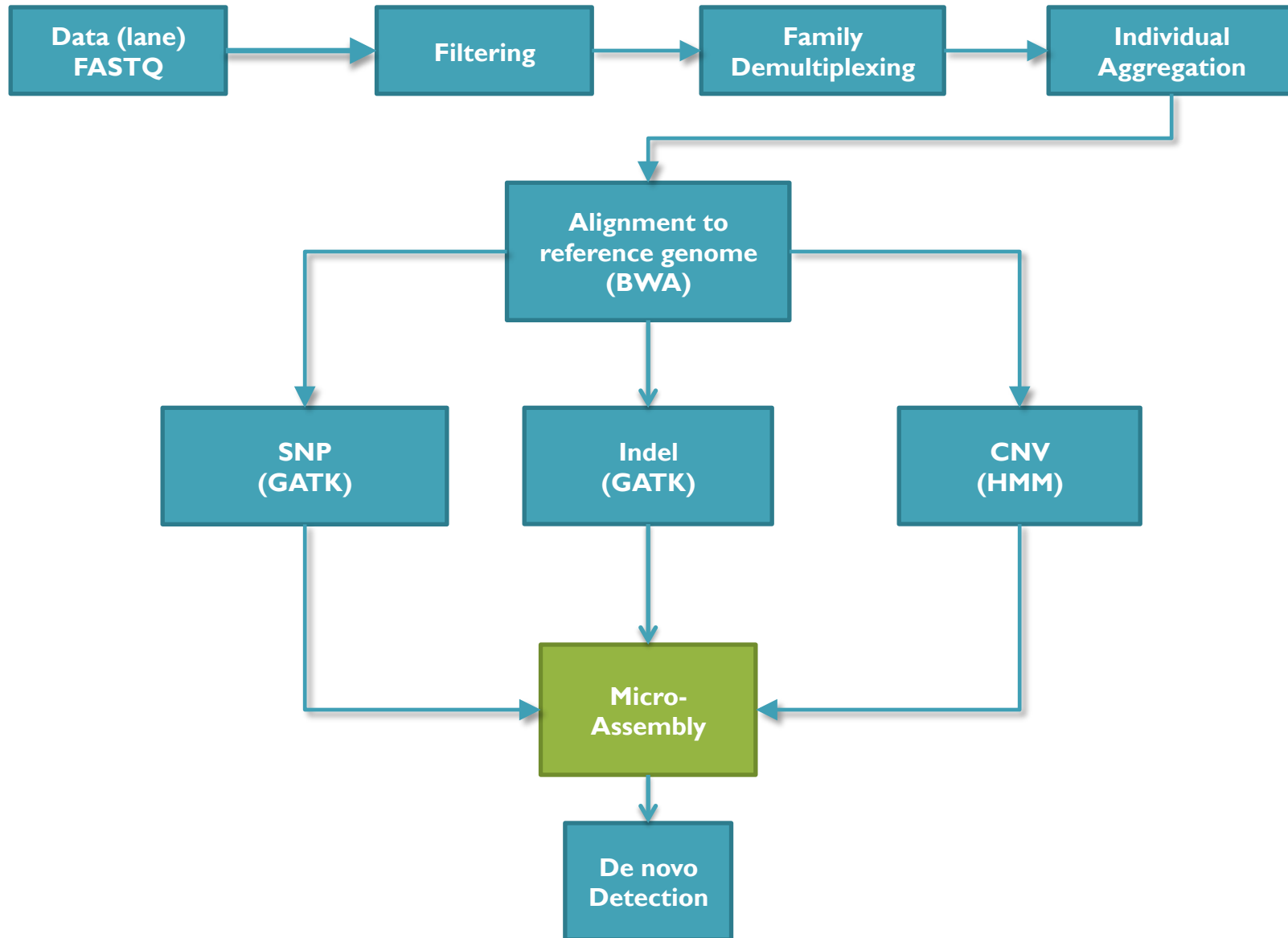
Families prepared and captured together to minimize batch effects

- Exome-capture performed with NimbleGen SeqCap EZ Exome v2.0 targeting 36 Mb of the genome.
- ~80% of the target at >20x coverage with ~93bp reads

**De novo gene disruptions in children on the autism spectrum**

lossifov *et al.* (2012) *Neuron*. 74:2 285-299

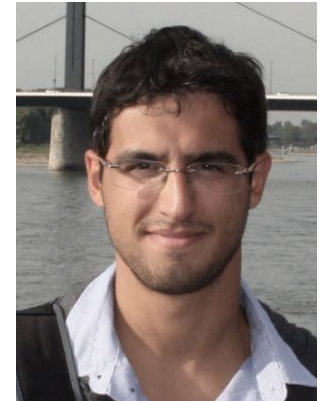
# Exome Sequencing Pipeline





# Scalpel: Haplotype Microassembly

G. Narzisi, D. Levy, I. Iossifov, J. Kendall, M. Wigler, M. Schatz



DNA sequence **micro-assembly** pipeline for accurate detection and validation of *de novo* mutations (SNPs, indels) within exome-capture data.

## Features

1. Combine **mapping** and **assembly**
2. Exhaustive search of **haplotypes**
3. **De novo mutations**

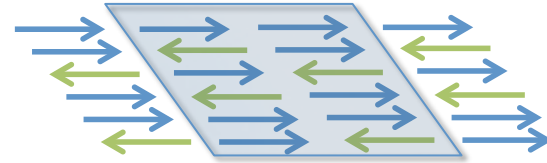


NRXN1 *de novo* SNP  
(auSSC12501 chr2:50724605)

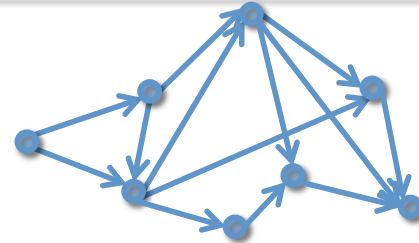


# Scalpel Pipeline

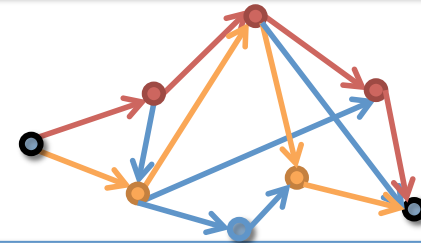
Extract reads mapping within the exon including (1) well-mapped reads, (2) soft-clipped reads, and (3) anchored pairs



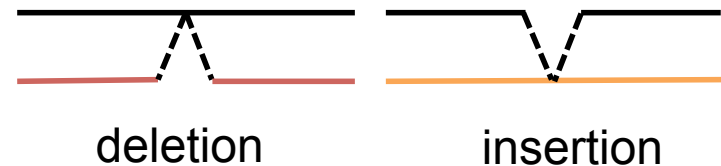
Decompose reads into overlapping  $k$ -mers and construct de Bruijn graph from the reads



Find end-to-end haplotype paths spanning the region



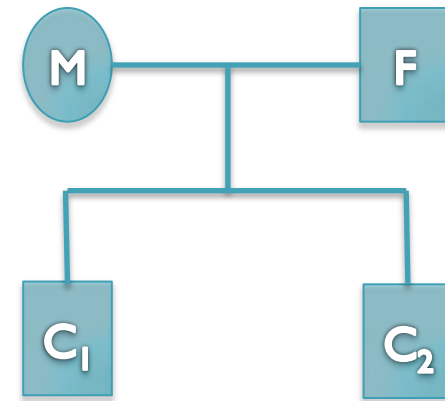
Align assembled sequences to reference to detect mutations



# De novo mutation discovery and validation

**Concept:** Identify mutations not present in parents.

**Challenge:** Sequencing errors in the child or low coverage in parents lead to false positive de novos



**Ref:** ...TCAGAACAGCTGGATGAGATCTTAGCCAACCTACCAGGAGATTGTCTTTGCCCGGA...

**Father:** ...TCAGAACAGCTGGATGAGATCTTAGCCAACCTACCAGGAGATTGTCTTTGCCCGGA...

**Mother:** ...TCAGAACAGCTGGATGAGATCTTAGCCAACCTACCAGGAGATTGTCTTTGCCCGGA...

**Sib:** ...TCAGAACAGCTGGATGAGATCTTAGCCAACCTACCAGGAGATTGTCTTTGCCCGGA...

**Aut(1):** ...TCAGAACAGCTGGATGAGATCTTAGCCAACCTACCAGGAGATTGTCTTTGCCCGGA...

**Aut(2):** ...TCAGAACAGCTGGATGAGATCTTACC-----CCGGGAGATTGTCTTTGCCCGGA...

6bp heterozygous deletion at chr13:25280526 ATP12A

# De novo Genetics of Autism

- In 343 family quads so far, we see significant enrichment in de novo **likely gene killers** in the autistic kids
  - Overall rate basically 1:1 (432:396)
  - 2:1 enrichment in nonsense mutations
  - 2:1 enrichment in frameshift indels
  - 4:1 enrichment in splice-site mutations
  - Most de novo originate in the paternal line in an age-dependent manner (56:18 of the mutations that we could determine)
- Observe strong overlap with the 842 genes known to be associated with fragile X protein FMR1
  - Related to neuron development and synaptic plasticity
  - Also strong overlap with chromatin remodelers

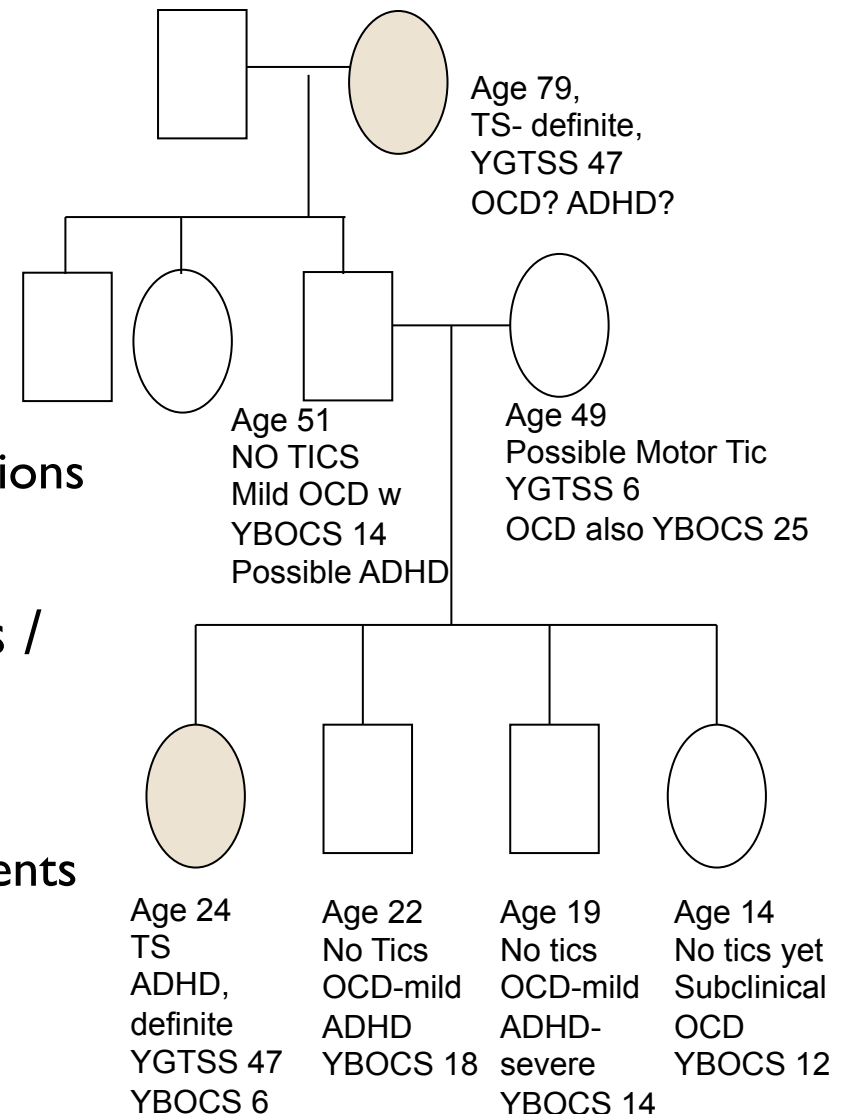
**De novo gene disruptions in children on the autism spectrum**

Iossifov *et al.* (2012) *Neuron*. 74:2 285-299

# Applications to ADHD & Tourette's

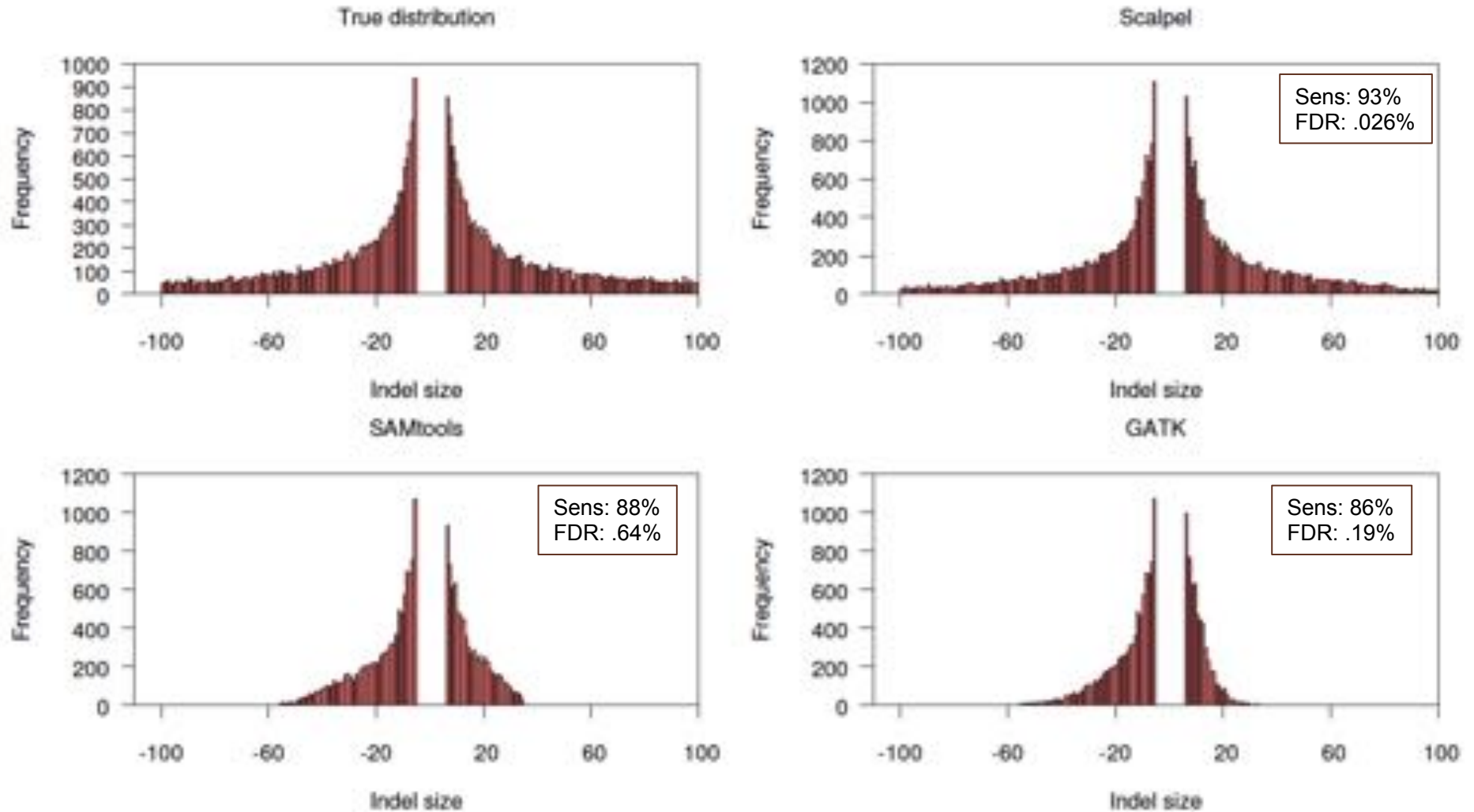
J. O'Rawe, G. Narzisi, M. Schatz, G. Lyon

- We believe similar mechanisms are involved in ADHD and Tourette's syndrome
  - Begun sequencing of families
  - Identify de novo and segregating mutations
- Cross analysis of GATK / SAMTools / SOAPindel / Scapel
  - High concordance on small events
  - Scalpel tends to identify more large events
  - Extensive wetlab validation in progress



# Scapel Indel Discovery

Indel size distribution (length > 5 bp)



**Detection of de novo mutations in exome-capture data using micro-assembly**  
Narzisi *et al.* (2012) *In preparation*

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2. **Plant Genome Assembly**
  1. Long read single molecule sequencing
  2. Other applications

# Genome Assembly Projects



## Sacred lotus

*Nelumbo nucifera* Gaertn.

Ming, R, et al. (2012) *Under Review*

Known for religious significance, herbal medicines, seed longevity, and water repellency

Illumina + 454 sequencing

- 900 Mbp Genome Size
- Low Heterozygosity

=> Excellent assembly



## Red Raspberry

*Rubus ideaus* L.

Price, J, et al. (2012) *In prep*

Member of the Rosacea family along with apple, pear, peach, strawberry.

Illumina + 454 sequencing

- 300 Mbp Genome Size
- High Heterozygosity

=> Good assembly



## Wheat DD

*Aegilops tauschii*

Schatz/Ware/McCombie collab.

One of the most important cereal crops in the world, one of three ancestral species of allohexaploid bread wheat

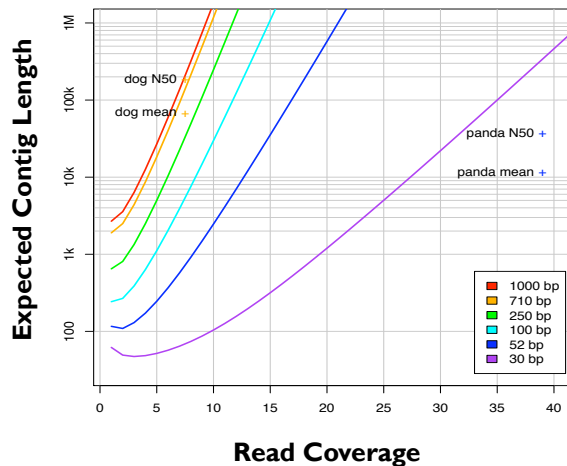
Illumina sequencing

- 4.5 Gbp Genome Size
- High repeat content

=> Challenged assembly

# Ingredients for a good assembly

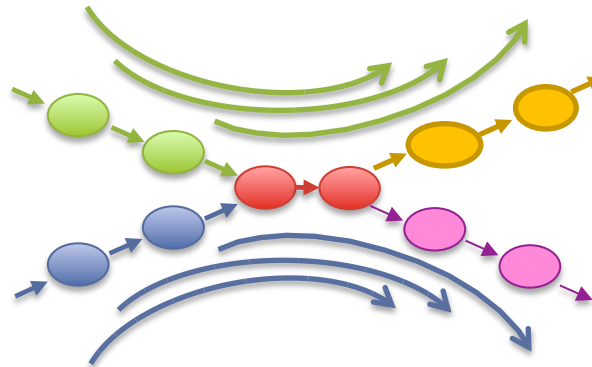
## Coverage



### High coverage is required

- Oversample the genome to ensure every base is sequenced with long overlaps between reads
- Biased coverage will also fragment assembly

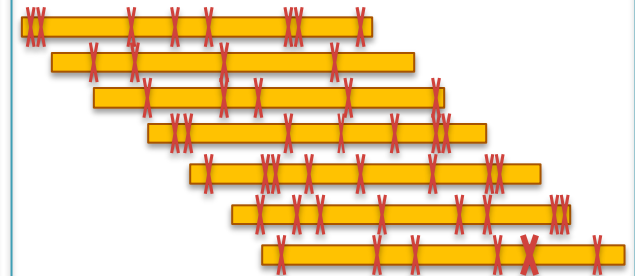
## Read Length



### Reads & mates must be longer than the repeats

- Short reads will have **false overlaps** forming hairball assembly graphs
- With long enough reads, assemble entire chromosomes into contigs

## Quality



### Errors obscure overlaps

- Reads are assembled by finding kmers shared in pair of reads
- High error rate requires very short seeds, increasing complexity and forming assembly hairballs

## Current challenges in *de novo* plant genome sequencing and assembly

Schatz MC, Witkowski, McCombie, WVR (2012) *Genome Biology*. 12:243



# Hybrid Sequencing



## **Illumina**

*Sequencing by Synthesis*

High throughput (60Gbp/day)

High accuracy (~99%)

Short reads (~100bp)



## **Pacific Biosciences**

*SMRT Sequencing*

Lower throughput (600Mbp/day)

Lower accuracy (~85%)

Long reads (2-5kbp+)



# PacBio Error Correction

<http://wgs-assembler.sf.net>

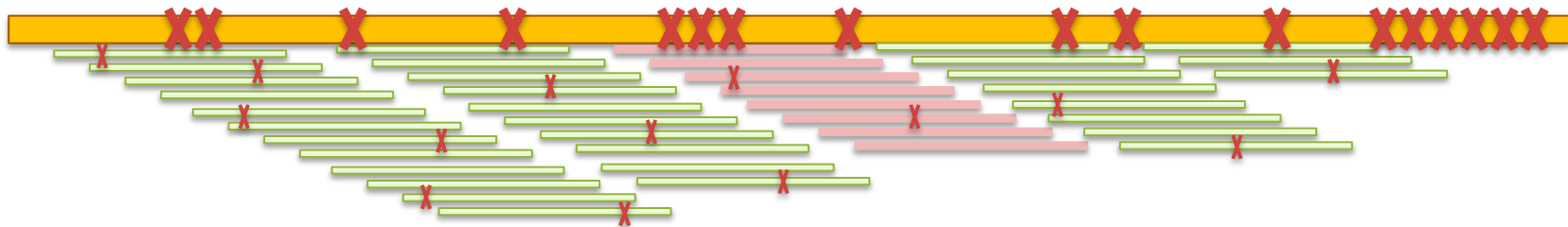


## I. Correction Pipeline

1. Map short reads (SR) to long reads (LR)
2. Trim LR at coverage gaps
3. Compute consensus for each LR

## 2. Error corrected reads can be easily assembled, aligned

1. Improves accuracy from ~85% to ~99%



**Hybrid error correction and de novo assembly of single-molecule sequencing reads.**

Koren, S, Schatz, MC, *et al.* (2012) *Nature Biotechnology*. doi:10.1038/nbt.2280

# SMRT-Assembly Results



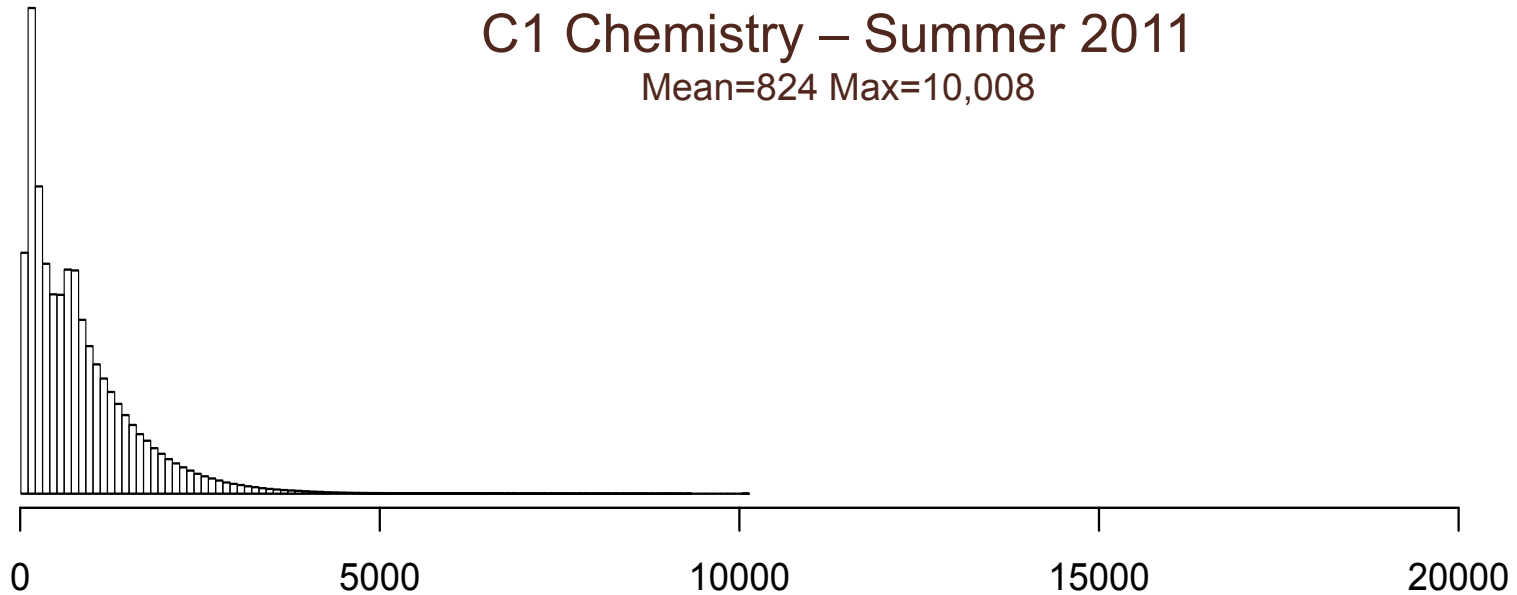
Organism	Technology	Reference bp	Assembly bp	# Contigs	Max Contig Length	N50
<i>Lambda</i> NEB3011 (median: 727 max: 3 280)	Illumina 100X 200bp	48 502	48 492	1	48 492 / 48 492	48 492 / 48 492 (100%) *
	PacBio PBcR 25X		48 440	1	48 444 / 48 444	48 444 / 48 440 (100%) *
<i>E. coli</i> K12 (median: 747 max: 3 068)	Illumina 100X 500bp	4 639 675	4 462 836	61	221 615 / 221 553	100 338 / 83 037 (82.36%) *
	PacBio PBcR 18X		4 465 533	77	239 058 / 238 224	71 479 / 68 309 (95.57%) *
	Both 18X PacBio PBcR + Illumina 50X 500bp		4 576 046	65	238 272 / 238 224	93 048 / 89 431 (96.11%) *
<i>E. coli</i> C227-11 (median: 1 217 max: 14 901)	PacBio CCS 50X	5 504 407	4 917 717	76	249 515	100 322
	PacBio 25X PBcR (corrected by 25X CCS)		5 207 946	80	357 234	98 774
	Both PacBio PBcR 25X + CCS 25X		5 269 158	39	647 362	227 302
	PacBio 50X PBcR (corrected by 50X CCS)		5 445 466	35	1 076 027	376 443
	Both PacBio PBcR 50X + CCS 25X		5 453 458	33	1 167 060	527 198
	Manually Corrected ALLORA Assembly <sup>8</sup>		5 452 251	23	653 382	402 041
<i>S. cerevisiae</i> S228c (median: 674 max: 5 994)	Illumina 100X 300bp	12 157 105	11 034 156	192	266 528 / 227 714	73 871 / 49 254 (66.68%) *
	PacBio PBcR 13X		11 110 420	224	224 478 / 217 704	62 898 / 54 633 (86.86%) *
	Both PacBio PBcR 13X + Illumina 50X 300bp		11 286 932	177	262 846 / 260 794	82 543 / 59 792 (72.44%) *
<i>Melospirinae asulanae</i> (median 997, max 13 079)	Illumina 194X (220/500/800 paired-end 2/5/10Kb mate-pairs)	1.23 Gbp	1 023 532 850	24 181	1 050 202	47 383
	454 15.4X (FLX + FLX Plus + 3/8/20Kbp paired-ends)		999 168 029	16 574	751 729	75 178
	454 15.4X + PacBio PBcR 3.75X		1 071 356 415	15 081	1 238 843	99 573

Hybrid assembly results using error corrected PacBio reads  
 Meets or beats Illumina-only or 454-only assembly in every case  
 \*\*\* Also useful for transcriptome and CNV analysis \*\*\*

# PacBio Long Read Sequencing

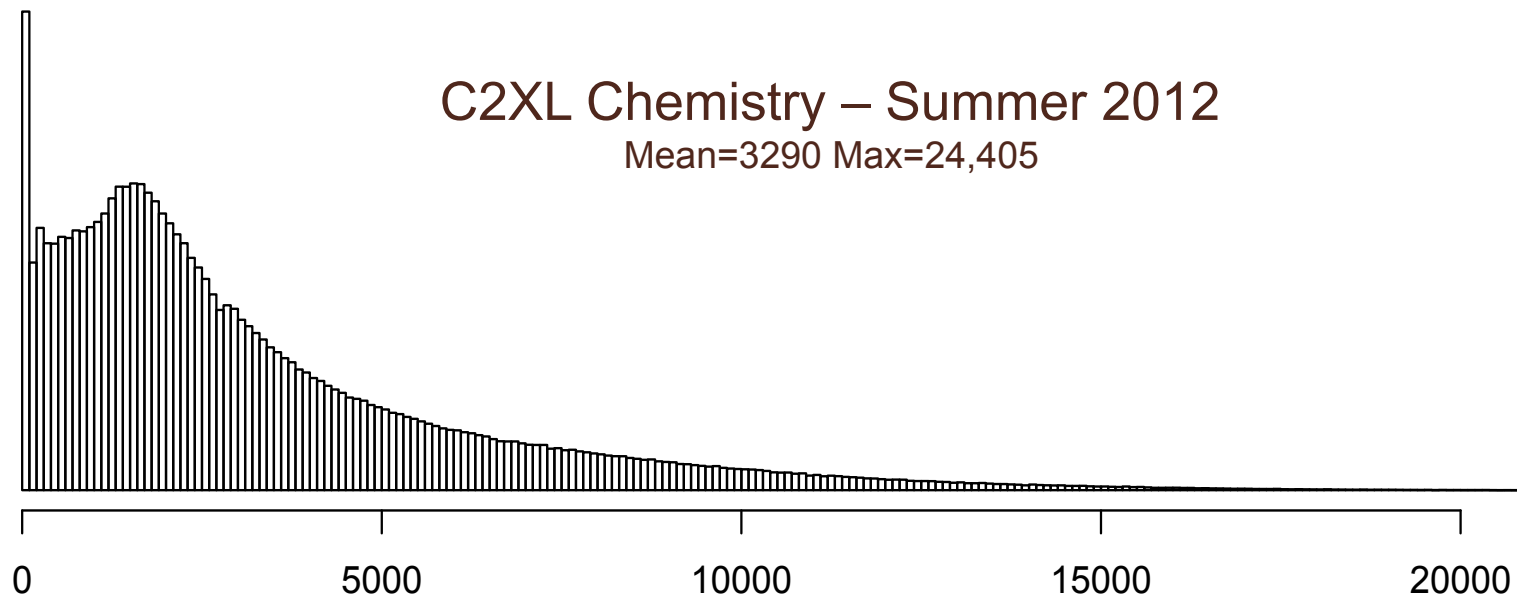
C1 Chemistry – Summer 2011

Mean=824 Max=10,008

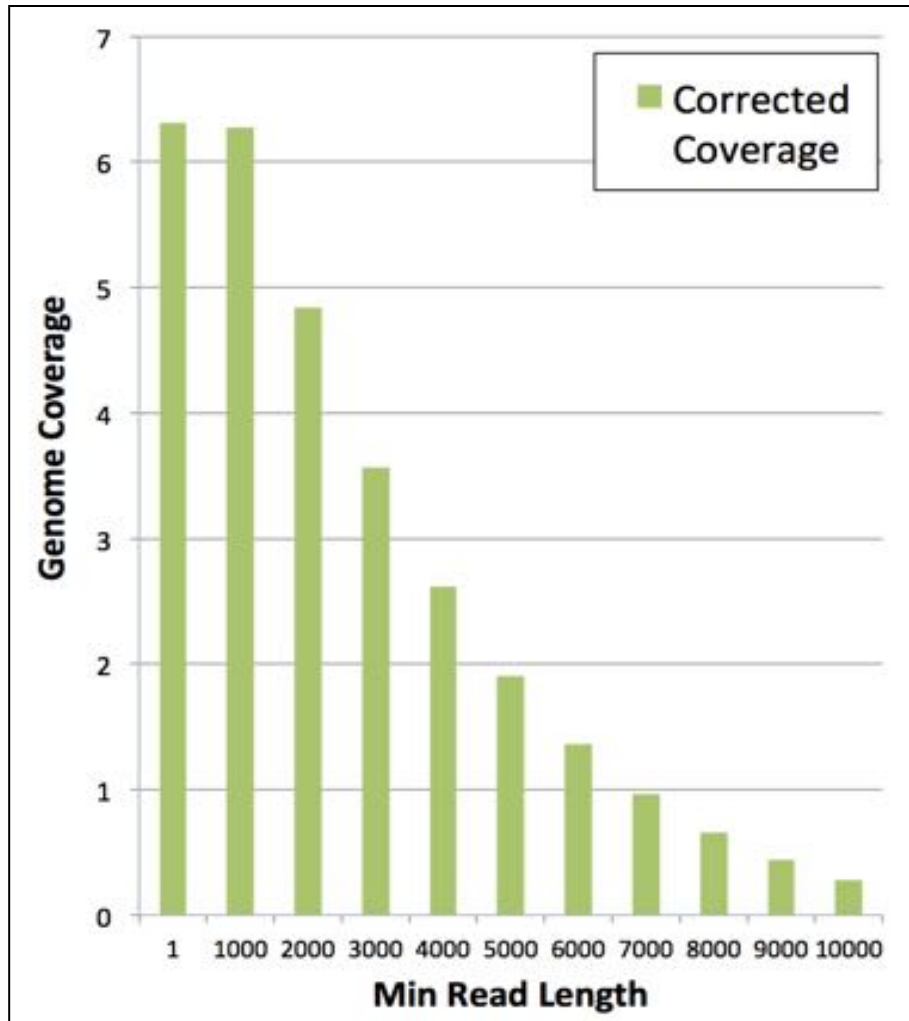


C2XL Chemistry – Summer 2012

Mean=3290 Max=24,405



# Preliminary Rice Assemblies



Assembly	Contig N50
Illumina Fragments 50x 2x100bp @ 180	3,925
MiSeq Fragments 23x 459bp 8x 2x251bp @ 450	6,444
PBeCR Reads 6.3x 2146bp ** MiSeq for correction	13,600
Illumina Mates 50x 2x100bp @ 180 36x 2x50bp @ 2100 51x 2x50bp @ 4800	13,696
PBeCR + Illumina Shred 6.3x 2146bp ** MiSeq for correction 51x 2x50bp @ 4800	25,108

In collaboration with McCombie & Ware labs @ CSHL

# Other Research Projects



High Performance  
Variant Detection  
And Interpretation

>168-fold speed up  
genotyping maize

## Answering the demands of digital genomics

Titmus, MA, Gurtowski, J, Schatz, MC (2012)

*Concurrency and Computation: Practice and Experience*

Analyzing  
Genomic Repeats and  
Sequencing Libraries

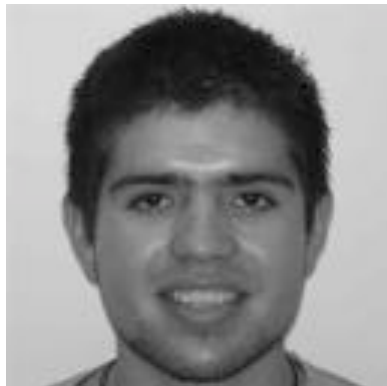
Pinpoint the regions  
we cant sequence with  
today's tech



## Genomic Dark Matter

Lee, H., Schatz, M.C. (2012)

*Bioinformatics. 28 (16): 2097-2105.*



Merge different  
assemblies into a high-  
accuracy consensus

Fix mistakes and capture  
all the information

## Improving Genome Assembly with Meta-assembly

Wences, A, Schatz, M.C. (2012)

*In preparation*

Evaluate the limits of  
assembling human, wheat  
and other genomes

How long is long  
enough?

## Assembly Complexity of Long Sequencing Reads

Marcus S, Lee, H., Schatz, M.C. (2012)

*In preparation*

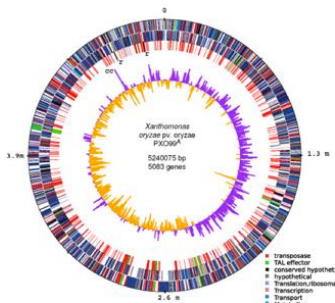


# Summary

I'm interested in answering biological questions by developing and applying novel algorithms and computational systems

- Interesting biological systems: human diseases, foods, biofuels
- Interesting biotechnology: new sequencing technologies
- Interesting computational systems: parallel & cloud technology
- Interesting algorithms: assembly, alignment, interpretation

Also extremely excited to teach the next generation of scientists in the WSBS, URP, and high school programs





# Acknowledgements

## Schatz Lab

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Deepak Nettem  
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Piyush Kansal  
Eric Biggers  
Aspyn Palatnick

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Hannon Lab  
Iossifov Lab  
Levy Lab  
Lippman Lab  
Lyon Lab  
Martienssen Lab  
McCombie Lab  
Ware Lab  
Wigler Lab

IT Department

## NBACC

Adam Phillippy  
Sergey Koren



# Thank You!

<http://schatzlab.cshl.edu/>  
[@mike\\_schatz](#)

